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Research Article

The lifespan of corpora lutea in nonpregnant females is positively correlated with gestation length

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Abstract

In mammals, a temporary endocrine gland called the corpus luteum forms on the ovary shortly after ovulation and is required for the initiation and maintenance of early pregnancy. However, the corpus luteum persists even when fertilization or pregnancy does not occur, and species-specifc variation in the length of this persistence remains enigmatic. Here we perform a comparative evolutionary study across 72 species and show that corpus luteum lifespan in nonpregnant females is positively correlated with gestation length. We argue that the most likely explanation for this correlation is physiological inertia. The corpus luteum begins secreting progesterone prior to implantation, and when pregnancy does not occur it takes time for females to degrade it and prepare the next reproductive cycle. Our study suggests that this physiological inertia is stronger in species with long gestation times.

Key words: corpus luteum, gestation, luteal phase, pregnancy.

Female mammals exhibit extensive maternal investment through gestation, lactation, and nursing of their offspring ([Clark and Poole](#page-4-0) [1967;](#page-4-0) [Gemmell 1981](#page-5-0); [Gilbert 1984](#page-5-1); [Van Raaij et al. 1989](#page-6-0); [Creel et](#page-5-2) [al. 1991](#page-5-2); [Urison and Buffenstein 1995](#page-6-1); [Prentice and Goldberg 2000;](#page-6-2) [Gobello 2021\)](#page-5-3). The corpus luteum, a temporary endocrine gland that develops from a ruptured ovarian follicle following ovulation, secretes progesterone and plays a critical role in the initiation and maintenance of early pregnancy in many mammalian species ([Moor 1968;](#page-5-4) [Sharman 1970](#page-6-3); [Tyndale-Biscoe et al. 1974;](#page-6-4) [Gadsby and](#page-5-5) [Landis Keyes 1984;](#page-5-5) [Harder et al. 1985;](#page-5-6) [Hinds 1990](#page-5-7); [Wiltbank 1994;](#page-6-5) [Bachelot and Binart 2005](#page-4-1); [McAllan 2011](#page-5-8); [Amelkina et al. 2015\)](#page-4-2). A functional corpus luteum shifts multiple physiological characteristics in pregnant females, including brain and blood chemistry, bone remodeling, mammary development, uterine development, and immune function ([Csapo and Wiest 1969](#page-5-9); [Graham and Clarke 1997;](#page-5-10) [Karlsson et al. 2001;](#page-5-11) [Neville et al. 2001](#page-5-12); [Sheehan and Numan 2002;](#page-6-6) [Conneely et al. 2003,](#page-4-3) [2008](#page-4-4); [Verstegen-Onclin and Verstegen 2008;](#page-6-7) [Fu and Levine 2009;](#page-5-13) [Brunton and Russell 2010](#page-4-5); [Mesiano et al. 2011;](#page-5-14) [Shah et al. 2018](#page-6-8); [Wu et al. 2018\)](#page-6-9).

Even when a reproductive cycle has not culminated in a pregnancy, the corpus luteum remains functional for varying lengths of time across species. In some cases, a functional corpus luteum can cause nonpregnant females to exhibit physiological changes similar to pregnancy ([Casida and Warwick 1945;](#page-4-6) [Weitlauf and Greenwald](#page-6-10) [1967;](#page-6-10) [Csapo and Wiest 1969](#page-5-9); [Csapo et al. 1972;](#page-5-15) [Shorey and Hughes](#page-6-11) [1973;](#page-6-11) [Csapo and Pulkkinen 1978;](#page-5-16) [Harder et al. 1985](#page-5-6); [Baird 1992](#page-4-7)). Indeed, in many species, a functional corpus luteum prevents a female from entering the next ovulatory cycle ([Tyndale-Biscoe et al.](#page-6-4)

[1974](#page-6-4); [Tyndale-Biscoe and Renfree 1987](#page-6-12)). What factors might explain variation in the persistence of functional corpora lutea in nonpregnant females?

It is possible that the same molecular mechanisms that maintain a functional corpus luteum in pregnancy may lead to their persistence even when pregnancy has not occurred. This hypothesis of physiological inertia predicts a positive correlation between gestation length and corpora lutea lifespan in nonpregnant females. In addition, monestrus species and seasonally breeding species are predicted to evolve mechanisms that reduce corpora lutea lifespans in nonpregnant females, because they must return to a new reproductive cycle or run the risk of missing their opportunity to reproduce. Here we compile data from 72 species from 12 orders and implement a variety of evolutionary methods to test these hypotheses.

Materials and methods. **Corpora lutea lifespan.**

We searched the literature for reports of functional corpora lutea in nonpregnant females across any species (without focusing on any region of the phylogeny). We considered a corpus luteum to be functional if histological data showed robust vascularization, or indirectly from the well-characterized spike in progesterone that corpora lutea produce in the early stages of implantation and pregnancy. We only included studies where females could not have been pregnant—that is, the study had to explicitly state that females were either housed separately from males or were housed with sterile (e.g., vasectomized) males. Otherwise, functional corpora

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lutea in "nonpregnant" females could have arisen if females were pregnant but aborted prior to birth. Our primary data came from searches in Google Scholar ([https://www.scholar.google.com\)](https://www.scholar.google.com) and Web of Science [\(https://webofscience.com/\)](https://webofscience.com/), with the phrases: *progesterone*, *atresia*, *corpus luteum*, *corpora lutea*, *period*, *corpus albicans*, *corpora albicantia*, *corpora albanica*, *menstruation*, *pseudopregnancy*, *nonpregnant*, *ruptured follicle*, *accessory corpus luteum*, and *corpus luteum regression*.

There are 2 main types of ovulation in mammals—spontaneous and induced. *Spontaneous ovulators* do not require any male cues or signals and will ovulate even in isolation from males, although male cues such as musk, urine, touch, and sounds can still infuence corpus luteum lifespan [\(Bruce 1960;](#page-4-8) [Bruce and Parrott 1960](#page-4-9); [DeMatteo et al. 2006](#page-5-17); [Roberts et al. 2012](#page-6-13)). For spontaneous ovulators, we recorded corpora lutea lifespan from any studies where females were not pregnant, including studies where females were isolated from males.

In contrast, *induced ovulators* require male cues to induce ovulation [\(Johnston et al. 2000](#page-5-18); [Brown 2011](#page-4-10); [Dixson 2021](#page-5-19)). In some cases, copulation or semen is required to induce ovulation ([Chen et al.](#page-4-11) [1985;](#page-4-11) [Johnston et al. 2004](#page-5-20); [Kauffman and Rissman 2006;](#page-5-21) [Ballantyne](#page-4-12) [2015;](#page-4-12) [Schjenken and Robertson 2020](#page-6-14)). One species (Djungarian Hamster, *Phodopus sungorus*) requires male cues at 2 different stages: one to ovulate, and a second exposure to induce the activation of corpora lutea ([Wynne-Edwards et al. 1987;](#page-6-15) [Erb et al. 1993;](#page-5-22) [Edwards](#page-5-23) [et al. 1994;](#page-5-23) [McMillan and Wynne-Edwards 1999\)](#page-5-24). For induced ovulators, we recorded corpora lutea lifespans from any studies where females were exposed to males but could not have been pregnant, e.g., studies where females were housed separately but exposed to male cues like scent, or studies where females were housed with vasectomized males.

Females of some species can undergo both spontaneous and induced ovulation [\(Hill and O'Donoghue 1913](#page-5-25); [Foresman and Daniel](#page-5-26) [1983;](#page-5-26) [Hinds 1989](#page-5-27); [Clamon Schulz et al. 2003](#page-4-13); [Steinman et al. 2012\)](#page-6-16). For these species, we only used data in the absence of males for evolutionary analyses.

We also scored corpora lutea lifespans from truly pregnant females, although data were scarcer than for nonpregnant females (*n* = 21 species; [Supplementary Data SD1\)](http://academic.oup.com/jmammal/article-lookup/doi/10.1093/jmammal/gyae114#supplementary-data). Progesterone production shifts from the corpus luteum to the placenta at some point during pregnancy, so we could not use progesterone levels to indicate a functional corpus luteum. Instead, we relied on robust histological analyses or ultrasound studies as a proxy for corpus luteum lifespan in pregnant females ([Supplementary Data SD1](http://academic.oup.com/jmammal/article-lookup/doi/10.1093/jmammal/gyae114#supplementary-data)).

Our exhaustive literature search yielded data from 72 species that fell into 6 monophyletic groups: metatherians (23 species from 4 orders); Order Carnivora (20 species); Order Rodentia + Order Lagomorpha (7 species); Order Artiodactyla + Order Perissodactyla (10 species); Order Primates (9 species); and Order Cingulata + Order Pilosa (3 species).

Gestation length and body size.

We searched the literature for gestation length data from the same species for which we gathered corpora luteum lifespan data. Because many physiological features of an organism correlate with body size, we also included body mass of adult females (nonpregnant) as a covariate in the evolutionary models. Most body mass data were derived from [Silva \(1995\),](#page-6-17) supplemented with additional literature searches [\(Supplementary Data SD1\)](http://academic.oup.com/jmammal/article-lookup/doi/10.1093/jmammal/gyae114#supplementary-data).

Phylogenetic generalized least squares.

To test for a correlation between corpus luteum lifespan and gestation length, we employed generalized least squares implemented in the GLS function from the R package "nlme" (Pinheiro et al. [2022\)](#page-6-18). To account for phylogenetic dependence, we included an expected correlation matrix derived from the mammalian phylogeny of [Upham et al. \(2019\),](#page-6-19) trimmed to only include the species in our data set [\(Supplementary Data SD2\)](http://academic.oup.com/jmammal/article-lookup/doi/10.1093/jmammal/gyae114#supplementary-data). We derived the correlation matrix with the corPAGEL function in the R package "ape" ([Paradis 2012\)](#page-6-20). Using R [\(R Core Team 2020\)](#page-6-21), the model we tested was GLS (corpus_luteum_lifespan \sim gestation_length + body_mass, correlation=corPagel, method="REML"), where "REML" indicates that the model was ft by maximizing the restricted log-likelihood, as advised by [Paradis \(2012\).](#page-6-20) All metrics were natural-logtransformed prior to regression.

Estrus frequency and breeding seasonality.

We hypothesized that monestrus species and seasonally breeding species would show relatively short corpora lutea lifespans in nonpregnant females. Therefore, we repeated the phylogenetic generalized least squares (PGLS) analyses above after subsetting out species for which we could collect estrus frequency and breeding seasonality.

Results

We scored corpora lutea lifespans in nonpregnant females, body weight of adult females, and gestation length from 72 species representing 12 orders. To our knowledge, this represents the most complete data set to date of species with data on all 3 traits ([Supplementary Data SD1\)](http://academic.oup.com/jmammal/article-lookup/doi/10.1093/jmammal/gyae114#supplementary-data). For a subset of 21 species, we found data on corpora lutea lifespans in pregnant females, as well as their frequency of ovulation and breeding seasonality [\(Supplementary Data](http://academic.oup.com/jmammal/article-lookup/doi/10.1093/jmammal/gyae114#supplementary-data) [SD1\)](http://academic.oup.com/jmammal/article-lookup/doi/10.1093/jmammal/gyae114#supplementary-data).

Corpora lutea lifespan in nonpregnant females was positively correlated with gestation length.

Corpus luteum lifespan in nonpregnant females was signifcantly positively correlated with gestation length $(F_{1,69} = 22.53, P = 0.0001)$, but not body mass $(F_{1,69} = 0.14, P = 0.70)$, with the full linear equation: corpus_luteum_lifespan = 0.78 × gestation_length − 0.02 × body_mass − 0.16. The lambda estimated from this full model was close to 1 (Lambda = 0.99), indicating that phylogenetic structure accounted for much of the relationships among variables.

Metatherians and Order Carnivora showed relatively long corpus luteum lifespan compared to the other 4 groups [\(Figs. 1](#page-2-0) and [2](#page-3-0)). Species with the largest gestational investment (ln(gestation) > 4.7) fell into 2 clusters: the 6 eutherian species that exhibit embryonic diapause showed relatively long-lived corpora lutea while almost all others showed relatively short-lived corpora lutea [\(Figs. 1](#page-2-0) and [2\)](#page-3-0). Embryonic diapause is a life history strategy in some mammalian species wherein embryos remain suspended at the blastocyst stage—sometimes for several months—before implanting in the uterus, all while the corpus luteum continues to produce progesterone [\(Hoffmann et al. 1978;](#page-5-28) [Foresman and Daniel 1983](#page-5-26); [Sempéré](#page-6-22) [et al. 1992](#page-6-22); [Sato et al. 2001](#page-6-23); [Clamon Schulz et al. 2003;](#page-4-13) [Okano et](#page-6-24) [al. 2006](#page-6-24); [Sattler and Polasek 2017](#page-6-25); [Larsen Tempel and Atkinson](#page-5-29) [2020\)](#page-5-29). The 6 eutherian species included in our analysis that exhibit embryonic diapause are *Ursus arctos* (Brown Bear), *U. americanus* (American Black Bear), *U. thibetanus* (Asian Black Bear), *Capreolus capreolus* (Siberian Roe Deer), *Eumetopias jubatus* (Steller Sea Lion), and *Odobenus rosmarus* (Walrus). The 3 bear species that exhibit embryonic diapause have notably longer corpus luteum lifespans than the fourth bear species included in our analysis, which does not exhibit embryonic diapause (Sun Bear, *Helarctos malayanus*; [Fig.](#page-3-0) [2\)](#page-3-0). Similarly, the only species in the Artiodactyla + Perissodactyla

Fig. 1. Corpus luteum lifespan in nonpregnant females was significantly positively correlated with gestation length. Each point on the plot is a species, placed into 1 of 6 groups indicated by color and image: red kangaroo represents 23 species of metatherians; black canid represents 20 species of Order Carnivora; green rodent represents 7 species of Order Rodentia + Order Lagomorpha; magenta deer represents 10 species of Order Artiodactyla + Order Perissodactyla; blue monkey represents 9 species of Order Primates; orange anteater represents 3 species of Order Cingulata + Order Pilosa. For both axes, the units are ln(days). Species images modifed from phylopic.org (exact fle names: PhyloPic.156b515d.Sarah-Werning.Callitrichoidea_Cebidae_ Cebinae_Platyrrhini.png; PhyloPic.570c7d9e.Alexandra-van-der-Geer.Rattus_Rattus-exulans.png; PhyloPic.6df900f7.Xavier-A-Jenkins.Myrmecophagidae_ pan-Myrmecophagidae_Tamandua_Tamandua-mexicana_Vermilingua.png; PhyloPic.96adba97.Margot-Michaud.Canis_Canis-simensis.png; PhyloPic.c306572a.Sarah-Werning.Macropus-Macropus.png; PhyloPic.cc03f5c2.Ferran-Sayol.Cervus-elaphus.png).

group that exhibits embryonic diapause (Roe Deer, *Capreolus capreolus*) shows a notable increase in corpus luteum lifespan compared to its relatives ([Fig. 2\)](#page-3-0).

Corpora lutea lifespan in *truly* **pregnant females was positively correlated with gestation length.**

Despite exhaustive literature searching, we were only able to score corpus luteum lifespan in truly pregnant females from 21 species (representing 8 orders; [Supplementary Data SD1](http://academic.oup.com/jmammal/article-lookup/doi/10.1093/jmammal/gyae114#supplementary-data)). Employing the same PGLS as described above, corpus luteum lifespan was positively correlated with gestation length $(F_{16} = 25.27, P = 0.0001)$ but not body size $(F_{16} = 1.52, P = 0.24)$ with the full linear equation: corpus_luteum_lifespan = $0.82 \times$ gestation_length + $0.08 \times$ body_mass $+ 0.67.$

Corpora lutea lifespan was relatively long in monestrus and seasonally breeding species.

From the 72 species in our full data set, we found data on estrus frequency and breeding seasonality for 51 and 47 species, respectively. Monestrus species (*n* = 9) had signifcantly longer corpora lutea lifespans in nonpregnant females than polyestrus species (*n* = 42), as indicated by signifcantly higher residuals from the full model above ($t = 2.4$, $df = 49$, $P = 0.02$). Seasonally breeding species ($n =$ 26) had signifcantly longer corpora lutea lifespans in nonpregnant females compared to species that breed year-round (*n* = 21, *t* = 2.58, df = 45, *P* = 0.01). Both results are opposite our predictions. In other words, species that seemingly have "more to lose" by maintaining active corpora lutea when not pregnant actually show longer corpora lutea lifespans.

Discussion

The reason for variation in lifespan of the corpora lutea remains relatively unknown in mammals, refecting the continued need for basic research on female pregnancy [\(Hayssen and Orr 2017](#page-5-30); [Hayssen 2020](#page-5-31)). During pregnancy, female mammals experience changes to their anatomy, physiology, neurobiology, hormone levels, immune system, vulnerability to predation, and even lifespan [\(Supplementary Data SD3](http://academic.oup.com/jmammal/article-lookup/doi/10.1093/jmammal/gyae114#supplementary-data)). Many of these shifts depend on a functional corpus luteum. In every species studied here, we found evidence that functional corpora lutea persist in nonpregnant females.

The hormonal signaling associated with maintaining a pregnancy begins prior to actual conception of offspring, during the estrus cycle. A functional corpus luteum begins secreting progesterone prior to implantation and is critical for early pregnancy. Thus, when a reproductive cycle does not yield successful pregnancy, the corpus luteum may simply represent physiological inertia toward a state of pregnancy. In other words, even when pregnancy fails it takes some time to reverse course and degrade the corpus luteum prior to entering the next reproductive cycle. Our study implies that this physiological inertia is stronger among species with long gestations.

The species studied here vary tremendously in the molecular mechanisms by which they maintain or degrade corpora lutea. Hormones related to corpora lutea are generally classifed into 3 categories: (i) luteotropic hormones that sustain corpus luteum development or maintenance; (ii) luteolytic hormones that lead to the degradation of the corpus luteum; and (iii) antiluteolytic hormones that prevent the degradation of the corpus luteum. All 3 of these hormonal classes have been found in eutherian species, while only a single luteolytic hormone has been described from metatherian species ([Supplementary Data SD3\)](http://academic.oup.com/jmammal/article-lookup/doi/10.1093/jmammal/gyae114#supplementary-data).

[Inbaraj et al. \(2021\)](#page-5-32) showed that 9 eutherian species (horses, cattle, humans, camels, rats, sheep, deer, cats, and dogs) showed species-specifc combinations of hormones involved in luteal regression. Similarly, [Chavan et al. \(2016\)](#page-4-14) showed that primates, horses, and rodents each had distinct hormonal mechanisms of luteotropic signaling.

CL lifespan residuals

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Fig. 2. The phylogenetic distribution of corpus luteum lifespan in nonpregnant females. Taxon colors and images as in [Fig. 1](#page-2-0). Branches colored according to the residuals of corpus luteum lifespan regressed onto gestation length + body mass. For context, most recent common ancestor of the species shown is roughly 157 million years ago. Image credits as in [Fig. 1](#page-2-0).

In contrast, only 1 luteolytic hormone (Prostaglandin 2α) has been shown to infuence corpora lutea lifespans in metatherians ([Renfree and Young 1979;](#page-6-26) [Tyndale-Biscoe and Renfree 1987;](#page-6-12) [Hinds](#page-5-7) [1990\)](#page-5-7). Furthermore, it has been argued that the corpus luteum plays a relatively minor role in maintaining pregnancy in marsupials, where gestational investment is much shorter than in eutherians ([Shorey and Hughes 1973;](#page-6-11) [Tyndale-Biscoe et al. 1974;](#page-6-4) [Tyndale-](#page-6-12)[Biscoe and Renfree 1987;](#page-6-12) [McAllan 2011\)](#page-5-8). Despite major differences in molecular regulation of and reliance on corpora lutea, the positive correlation between corpora lutea lifespans in nonpregnant females and gestation length extends across mammals, including marsupials ([Figs. 1](#page-2-0) and [2\)](#page-3-0).

Without more comparative biology on the molecular pathways involved in corpora lutea, the positive correlation between their lifespan in nonpregnant females and gestation length will remain enigmatic. But the correlation implies the same pathways involved in maintaining a pregnancy also extend corpora lutea lifespans in females even when there is no pregnancy to support. This constraint may be especially strong in species with embryonic diapause because they must maintain an active corpus luteum for much longer than species that do not manifest embryonic diapause [\(Hoffmann et al. 1978](#page-5-28); [Foresman and Daniel 1983;](#page-5-26) [Tsubota et al. 1987](#page-6-27); [Sato et al. 2001;](#page-6-23) [Okano et al. 2006](#page-6-24); [Frederick](#page-5-33) [et al. 2010\)](#page-5-33). Our study offers an explanation for the persistence of functional corpora lutea in nonpregnant females, suggesting that it is constrained by the physiological preparation for a real pregnancy.

Supplementary data

Supplementary data are available at *Journal of Mammalogy* online.

Supplementary Data SD1. Spreadsheet containing all data analyzed, including literature sources.

Supplementary Data SD2. The phylogenetic tree used in the full PGLS.

Supplementary Data SD3. Citations to pregnancy-related changes and hormonal changes in females.

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Author contributions

Conceptualization: ML-S and MDD; Data curation: ML-S and CRG; Formal analysis: MDD; Funding acquisition: MDD; Investigation: ML-S; Methodology: ML-S, MDD, and CRG; Project administration: ML-S and MDD; Resources: MDD; Software: MDD; Supervision: MDD; Validation: MDD; Visualization: MDD; Writing—original draft: ML-S; Writing—review & editing: ML-S, MDD, and CRG.

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Confict of interest

None declared.

Data availability

Found in [Supplementary Data SD1.](http://academic.oup.com/jmammal/article-lookup/doi/10.1093/jmammal/gyae114#supplementary-data)

References

- [Amelkina O, Braun BC, Dehnhard M, Jewgenow K.](#page-0-1) 2015. The corpus luteum of the domestic cat: histologic classifcation and intraluteal hormone profle. Theriogenology 83(4):711–720. [https://doi.](https://doi.org/10.1016/j.theriogenology.2014.11.008) [org/10.1016/j.theriogenology.2014.11.008](https://doi.org/10.1016/j.theriogenology.2014.11.008)
- [Bachelot A, Binart N.](#page-0-2) 2005. Corpus luteum development: lessons from genetic models in mice. *Current Topics in Developmental Biology* 68:49–84. [https://doi.org/10.1016/S0070-2153\(05\)68003](https://doi.org/10.1016/S0070-2153(05)68003)
- [Baird DT.](#page-0-3) 1992. Luteotrophic control of the corpus luteum. Animal Reproduction Science 28(1–4):95–102. [https://doi.](https://doi.org/10.1016/0378-4320(92)90096-v) [org/10.1016/0378-4320\(92\)90096-v](https://doi.org/10.1016/0378-4320(92)90096-v)
- [Ballantyne K.](#page-1-0) 2015. New insights into the reproductive physiology and management of the female koala (*Phascolarctos cinereus*): factors affecting the control of the oestrous cycle [PhD dissertation]. [Brisbane (Australia)]: The University of Queensland.
- [Brown JL.](#page-1-1) 2011. Female reproductive cycles of wild female felids. Animal Reproduction Science 124(3–4):155–162. [https://doi.](https://doi.org/10.1016/j.anireprosci.2010.08.024) [org/10.1016/j.anireprosci.2010.08.024](https://doi.org/10.1016/j.anireprosci.2010.08.024)
- [Bruce HM.](#page-1-2) 1960. A block to pregnancy in the mouse caused by proximity of strange males. Journal of Reproduction and Fertility 1(1):96– 103.<https://doi.org/10.1530/jrf.0.0010096>
- [Bruce HM, Parrott D.](#page-1-3) 1960. Role of olfactory sense in pregnancy block by strange males. Science 131(3412):1526–1526. [https://doi.](https://doi.org/10.1126/science.131.3412.1526) [org/10.1126/science.131.3412.1526](https://doi.org/10.1126/science.131.3412.1526)
- [Brunton PJ, Russell JA.](#page-0-4) 2010. Endocrine induced changes in brain function during pregnancy. Brain Research 1364:198–215. [https://doi.](https://doi.org/10.1016/j.brainres.2010.09.062) [org/10.1016/j.brainres.2010.09.062](https://doi.org/10.1016/j.brainres.2010.09.062)
- [Casida LE, Warwick EJ.](#page-0-5) 1945. The necessity of the corpus luteum for maintenance of pregnancy in the ewe. Journal of Animal Science 4(1):34–36.<https://doi.org/10.2527/jas1945.4134>
- [Chavan AR, Bhullar B-AS, Wagner GP.](#page-2-1) 2016. What was the ancestral function of decidual stromal cells? A model for the evolution of eutherian pregnancy. Placenta 40:40–51. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.placenta.2016.02.012) [placenta.2016.02.012](https://doi.org/10.1016/j.placenta.2016.02.012)
- [Chen B, Yuen Z, Pan G.](#page-1-4) 1985. Semen-induced ovulation in the Bactrian camel (*Camelus bactrianus*). Reproduction 74(2):335–339. [https://](https://doi.org//10.1530/jrf.0.0740335) [doi.org/10.1530/jrf.0.0740335](https://doi.org//10.1530/jrf.0.0740335)
- [Clamon Schulz L, Nelson RA, Pyter LM, Bahr JM.](#page-1-5) 2003. Induction of pseudopregnancy in the American black bear (*Ursus americanus*). Journal of Experimental Zoology, Part A: Comparative Experimental Biology 298(2):162–166. [https://doi.org/10.1002/](https://doi.org/10.1002/jez.a.10269) [jez.a.10269](https://doi.org/10.1002/jez.a.10269)
- [Clark M, Poole W.](#page-0-6) 1967. The reproductive system and embryonic diapause in the female kangaroo, *Marcopodus giganteus*. Australian Journal of Zoology 15(3):441–459. [https://doi.org/10.1071/](https://doi.org/10.1071/zo9670441) [zo9670441](https://doi.org/10.1071/zo9670441)
- [Conneely OM, Jericevic BM, Arnett-Mansfeld R.](#page-0-7) 2008. Progesterone signaling in mammary gland development. In: Conneely O, Otto C, editors. Ernst Schering Foundation symposium proceedings, vol. 1. Berlin (Heidelberg): Springer Berlin Heidelberg; p. 175–185.
- [Conneely OM, Jericevic BM, Lydon JP.](#page-0-8) 2003. Progesterone receptors in mammary gland development and tumorigenesis. Journal of Mammary Gland Biology and Neoplasia 8(2):205–214. [https://doi.](https://doi.org/10.1023/a:1025952924864) [org/10.1023/a:1025952924864](https://doi.org/10.1023/a:1025952924864)
- [Creel SR, Monfort SL, Wildt DE, Waser PM.](#page-0-9) 1991. Spontaneous lactation is an adaptive result of pseudopregnancy. Nature 351(6328):660– 662. <https://doi.org/10.1038/351660a0>
- [Csapo AI, Pulkkinen M.](#page-0-10) 1978. Indispensability of the human corpus luteum in the maintenance of early pregnancy luteectomy evidence. Obstetrical & Gynecological Survey 33(2):69–81. [https://doi.](https://doi.org/10.1097/00006254-197802000-00001) [org/10.1097/00006254-197802000-00001](https://doi.org/10.1097/00006254-197802000-00001)
- [Csapo AI, Pulkkinen MO, Ruttner B, Sauvage JP, Wiest WG.](#page-0-11) 1972. The signifcance of the human corpus luteum in pregnancy maintenance: I. Preliminary studies. American Journal of Obstetrics and Gynecology 112(8):1061–1067. [https://doi.](https://doi.org/10.1016/0002-9378(72)90181-0) [org/10.1016/0002-9378\(72\)90181-0](https://doi.org/10.1016/0002-9378(72)90181-0)
- [Csapo AI, Wiest WG.](#page-0-12) 1969. An examination of the quantitative relationship between progesterone and the maintenance of pregnancy. Endocrinology 85(4):735–746. [https://doi.org/10.1210/](https://doi.org/10.1210/endo-85-4-735) [endo-85-4-735](https://doi.org/10.1210/endo-85-4-735)
- [DeMatteo KE, Porton IJ, Kleiman DG, Asa CS.](#page-1-6) 2006. The effect of the male bush dog (*Speothos venaticus*) on the female reproductive cycle. Journal of Mammalogy 87(4):723–732. [https://doi.](https://doi.org/10.1644/05-mamm-a-342r1.1) [org/10.1644/05-mamm-a-342r1.1](https://doi.org/10.1644/05-mamm-a-342r1.1)
- [Dixson AL.](#page-1-7) 2021. The evolution of mating-induced and spontaneous ovulation. In: Dixson AF, editor. Mammalian sexuality: the act of mating and the evolution of reproduction. Cambridge: Cambridge University Press; p. 248–264.
- [Edwards HE, Jenkins KL, Mucklow LC, Erb GE, Wynne-Edwards KE.](#page-1-8) 1994. Endocrinology of the pregnant Djungarian hamster *Phodopus campbelli*. Journal of Reproduction and Fertility 101(1):1–8. [https://](https://doi.org/10.1530/jrf.0.1010001) doi.org/10.1530/jrf.0.1010001
- [Erb G, Edwards H, Jenkins K, Mucklow L, Wynne-Edwards K.](#page-1-9) 1993. Induced components in the spontaneous ovulatory cycle of the Djungarian hamster (*Phodopus campbelli*). Physiology & Behavior 54(5):955–959. [https://doi.org/10.1016/0031-9384\(93\)90308-3](https://doi.org/10.1016/0031-9384(93)90308-3)
- [Foresman K, Daniel J.](#page-4-15) 1983. Plasma progesterone concentrations in pregnant and nonpregnant black bears (*Ursus americanus*). Reproduction 68(1):235–239. [https://doi.org/10.1530/](https://doi.org//10.1530/jrf.0.0680235) [jrf.0.0680235](https://doi.org//10.1530/jrf.0.0680235)
- [Frederick C, Kyes R, Hunt K, Collins D, Durrant B, Wasser SK.](#page-4-16) 2010. Methods of estrus detection and correlates of the reproductive cycle in the Sun Bear (*Helarctos malayanus*). Theriogenology 74(7):1121– 1135.<https://doi.org/10.1016/j.theriogenology.2010.05.010>
- [Fu Q, Levine BD.](#page-0-13) 2009. Autonomic circulatory control during pregnancy in humans. Seminars in Reproductive Medicine 27(4):330–337. <https://doi.org/10.1055/s-0029-1225261>
- [Gadsby JE, Landis Keyes P.](#page-0-14) 1984. Control of corpus luteum function in the pregnant rabbit: role of the placenta ("placental luteotropin") in regulating responsiveness of corpora lutea to estrogen1. Biology of Reproduction 31(1):16–24. [https://doi.org/10.1095/](https://doi.org/10.1095/biolreprod31.1.16) [biolreprod31.1.16](https://doi.org/10.1095/biolreprod31.1.16)
- [Gemmell R.](#page-0-15) 1981. The role of the corpus luteum of lactation in the bandicoot *Isoodon macrourus* (Marsupialia: Peramelidae). General and Comparative Endocrinology 44(1):13–19. [https://doi.](https://doi.org/10.1016/0016-6480(81)90350-6) [org/10.1016/0016-6480\(81\)90350-6](https://doi.org/10.1016/0016-6480(81)90350-6)
- [Gilbert AN.](#page-0-16) 1984. Postpartum and lactational estrus: a comparative analysis in Rodentia. Journal of Comparative Psychology 98(3):232–245. [https://doi.org/10.1037/0735-7036.98.3.232](https://doi.org//10.1037/0735-7036.98.3.232)
- [Gobello C.](#page-0-17) 2021. Revisiting canine pseudocyesis. Theriogenology 167:94–98.<https://doi.org/10.1016/j.theriogenology.2021.03.014>
- [Graham JD, Clarke CL.](#page-0-18) 1997. Physiological action of progesterone in target tissues*. Endocrine Reviews 18(4):502–519. [https://doi.](https://doi.org/10.1210/edrv.18.4.0308) [org/10.1210/edrv.18.4.0308](https://doi.org/10.1210/edrv.18.4.0308)
- [Harder J, Hinds L, Horn C, Tyndale-Biscoe C.](#page-0-19) 1985. Effects of removal in late pregnancy of the corpus luteum, graafan follicle or ovaries on plasma progesterone, oestradiol, LH, parturition and post-partum

oestrus in the tammar wallaby, *Macropus eugenii*. Reproduction 75(2):449–459. [https://doi.org/10.1530/jrf.0.0750449](https://doi.org//10.1530/jrf.0.0750449)

- [Hayssen V.](#page-2-2) 2020. Misconceptions about conception and other fallacies: historical bias in reproductive biology. Integrative and Comparative Biology 60(3):683–691. [https://doi.org/10.1093/icb/](https://doi.org/10.1093/icb/icaa035) [icaa035](https://doi.org/10.1093/icb/icaa035)
- [Hayssen V, Orr TJ.](#page-2-3) 2017. Reproduction in mammals: the female perspective. Baltimore (MD, USA): John Hopkins University Press.
- [Hill J, O'Donoghue CH.](#page-1-10) 1913. The reproductive cycle in the marsupial *Dasyurus viverrinus*. Journal of Cell Science 2(233):133–174. [https://](https://doi.org//10.1242/jcs.s2-59.233.133) [doi.org/10.1242/jcs.s2-59.233.133](https://doi.org//10.1242/jcs.s2-59.233.133)
- [Hinds L.](#page-4-17) 1990. Control of pregnancy, parturition and luteolysis in marsupials. Reproduction Fertility and Development 2(5):535–552. <https://doi.org/10.1071/rd9900535>
- [Hinds LA.](#page-1-11) 1989. Plasma progesterone through pregnancy and the estrous cycle in the eastern quoll, *Dasyurus viverrinus*. General and Comparative Endocrinology 75(1):110–117. [https://doi.](https://doi.org/10.1016/0016-6480(89)90015-4) [org/10.1016/0016-6480\(89\)90015-4](https://doi.org/10.1016/0016-6480(89)90015-4)
- [Hoffmann B, Barth D, Karg H.](#page-4-18) 1978. Progesterone and estrogen levels in peripheral plasma of the pregnant and nonpregnant roe deer (*Capreolus capreolus*). Biology of Reproduction 19(5):931–935. <https://doi.org/10.1095/biolreprod19.5.931>
- [Inbaraj KC, Yu Z, Chai S, Alagamuthu KK, Vallavan R, Meyyazhagan A,](#page-2-4) [Balasubramanian B, Chinappan G, Xu S, Yang G.](#page-2-4) 2021. Regression of corpus luteum in cetaceans: a systematic review. Zoology 149:125960. <https://doi.org/10.1016/j.zool.2021.125960>
- [Johnston S, McGowan M, O'Callaghan P, Cox R, Nicolson V.](#page-1-12) 2000. Studies of the oestrous cycle, oestrus and pregnancy in the koala (*Phascolarctos cinereus*). Reproduction 120(1):49–57. [https://doi.](https://doi.org/10.1530/reprod/120.1.49) [org/10.1530/reprod/120.1.49](https://doi.org/10.1530/reprod/120.1.49)
- [Johnston S, O'Callaghan P, Nilsson K, Tzipori G, Curlewis J.](#page-1-13) 2004. Semeninduced luteal phase and identifcation of a LH surge in the koala (*Phascolarctos cinereus*). Reproduction 128(5):629–634. [https://doi.](https://doi.org//10.1530/rep.1.00300) [org/10.1530/rep.1.00300](https://doi.org//10.1530/rep.1.00300)
- [Karlsson C, Obrant KJ, Karlsson M.](#page-0-20) 2001. Pregnancy and lactation confer reversible bone loss in humans. Osteoporosis International 12(10):828–834.<https://doi.org/10.1007/s001980170033>
- [Kauffman AS, Rissman EF.](#page-1-14) 2006. Neuroendocrine control of matinginduced ovulation. In: Knobil and Neill's physiology of reproduction. London: Academic Press; p. 2283–2326.
- [Larsen Tempel JT, Atkinson S.](#page-1-15) 2020. Endocrine profling of reproductive status and evidence of pseudopregnancy in the Pacifc walrus (*Odobenus rosmarus divergens*). PLoS One 15(9):e0239218. [https://](https://doi.org/10.1371/journal.pone.0239218) doi.org/10.1371/journal.pone.0239218
- [McAllan BM.](#page-4-19) 2011. Chapter 10 - reproductive endocrinology of prototherians and metatherians. In: Norris DO, Lopez KH, editors. Hormones and reproduction of vertebrates. London: Academic Press; p. 195–214.
- [McMillan HJ, Wynne-Edwards KE.](#page-1-16) 1999. Divergent reproductive endocrinology of the estrous cycle and pregnancy in dwarf hamsters (*Phodopus*). Comparative Biochemistry and Physiology, Part A: Molecular & Integrative Physiology 124(1):53–67. [https://doi.](https://doi.org/10.1016/s1095-6433(99)00090-2) [org/10.1016/s1095-6433\(99\)00090-2](https://doi.org/10.1016/s1095-6433(99)00090-2)
- [Mesiano S, Wang Y, Norwitz ER.](#page-0-21) 2011. Progesterone receptors in the human pregnancy uterus: do they hold the key to birth timing? Reproductive Sciences 18(1):6–19. [https://doi.](https://doi.org/10.1177/1933719110382922) [org/10.1177/1933719110382922](https://doi.org/10.1177/1933719110382922)
- [Moor RM.](#page-0-22) 1968. Effect of embryo on corpus luteum function. Journal of Animal Science 27(suppl 1):97–118. [https://doi.org/10.2527/](https://doi.org//10.2527/animalsci1968.27Supplement_197x) [animalsci1968.27Supplement_197x](https://doi.org//10.2527/animalsci1968.27Supplement_197x)
- [Neville MC, Morton J, Umemura S.](#page-0-23) 2001. Lactogenesis: the transition from pregnancy to lactation. Pediatric Clinics of North America 48(1):35–52. [https://doi.org/10.1016/s0031-3955\(05\)70284-4](https://doi.org/10.1016/s0031-3955(05)70284-4)
- [Okano T, Nakamura S, Nakashita R, Komatsu T, Murase T, Asano M,](#page-4-20) [Tsubota T.](#page-4-20) 2006. Incidence of ovulation without coital stimuli in captive Japanese black bears (*Ursus thibetanus japonicus*) based on serum progesterone profles. The Journal of Veterinary Medical Science 68(10):1133–1137. <https://doi.org/10.1292/jvms.68.11CC33>
- [Paradis E.](#page-1-17) 2012. Analysis of phylogenetics and evolution with R. New York (NY, USA): Springer.
- [Pinheiro J, Bates D, Team RC.](#page-1-18) 2022. nlme: linear and nonlinear mixed effects models. R package version 3.1-157. [https://CRAN.R-project.](https://CRAN.R-project.org/package=nlme) [org/package=nlme.](https://CRAN.R-project.org/package=nlme)
- [Prentice AM, Goldberg GR.](#page-0-24) 2000. Energy adaptations in human pregnancy: limits and long-term consequences. The American Journal of Clinical Nutrition 71(5 Suppl):1226S–1232S. [https://doi.](https://doi.org/10.1093/ajcn/71.5.1226s) [org/10.1093/ajcn/71.5.1226s](https://doi.org/10.1093/ajcn/71.5.1226s)
- [R Core Team](#page-1-19). 2020. R: a language and environment for statistical computing. Vienna (Austria): R Foundation for Statistical Computing. [http://www.R-project.org/.](http://www.R-project.org/)
- [Renfree MB, Young IR.](#page-4-21) 1979. Steroids in pregnancy and parturition in the marsupial, *Macropus eugenii*. Proceedings of the Fifth International Congress on Hormonal Steroids; p. 515–522.
- [Roberts EK, Lu A, Bergman TJ, Beehner JC.](#page-1-20) 2012. A Bruce effect in wild geladas. Science 335(6073):1222–1225. [https://doi.org/10.1126/](https://doi.org/10.1126/science.1213600) [science.1213600](https://doi.org/10.1126/science.1213600)
- [Sato M, Tsubota T, Komatsu T, Watanabe G, Taya K, Murase T, Kita I,](#page-4-22) [Kudo T.](#page-4-22) 2001. Changes in sex steroids, gonadotropins, prolactin, and inhibin in pregnant and nonpregnant Japanese black bears (*Ursus thibetanus japonicus*). Biology of Reproduction 65(4):1006– 1013. <https://doi.org/10.1095/biolreprod65.4.1006>
- [Sattler R, Polasek L.](#page-1-21) 2017. Serum estradiol and progesterone profles during estrus, pseudopregnancy, and active gestation in Steller sea lions. Zoo Biology 36(5):323–331.<https://doi.org/10.1002/zoo.21381>
- [Schjenken JE, Robertson SA.](#page-1-22) 2020. The female response to seminal fuid. Physiological Reviews 100(3):1077–1117. [https://doi.org/10.1152/](https://doi.org/10.1152/physrev.00013.2018) [physrev.00013.2018](https://doi.org/10.1152/physrev.00013.2018)
- [Sempéré AJ, Mauget R, Chemineau P.](#page-1-23) 1992. Experimental induction of luteal cyclicity in roe deer (*Capreolus capreolus*). Journal of Reproduction and Fertility 96(1):379–384. [https://doi.org/10.1530/](https://doi.org/10.1530/jrf.0.0960379) [jrf.0.0960379](https://doi.org/10.1530/jrf.0.0960379)
- [Shah NM, Imami N, Johnson MR.](#page-0-25) 2018. Progesterone modulation of pregnancy-related immune responses. Frontiers in Immunology 9:1293. [https://doi.org/10.3389/fmmu.2018.01293](https://doi.org/10.3389/fimmu.2018.01293)
- [Sharman G.](#page-0-26) 1970. Reproductive physiology of marsupials. Science 167(3922):1221–1228.<https://doi.org/10.1126/science.167.3922.12>
- [Sheehan T, Numan M.](#page-0-27) 2002. Estrogen, progesterone, and pregnancy termination alter neural activity in brain regions that control maternal behavior in rats. Neuroendocrinology 75(1):12–23. [https://doi.](https://doi.org/10.1159/000048217) [org/10.1159/000048217](https://doi.org/10.1159/000048217)
- [Shorey C, Hughes R.](#page-4-23) 1973. Developement, function and regression of the corpus luteum in the marsupial *Trichosurus vulpecula*. Australian Journal of Zoology 21(4):477–489.<https://doi.org/10.1071/zo9730477>
- [Silva M.](#page-1-24) 1995. CRC handbook of mammalian body masses. Boca Raton (FL, USA): CRC Press.
- [Steinman KJ, O'Brien JK, Monfort SL, Robeck TR.](#page-1-25) 2012. Characterization of the estrous cycle in female beluga (*Delphinapterus leucas*) using urinary endocrine monitoring and transabdominal ultrasound: evidence of facultative induced ovulation. General and Comparative Endocrinology 175(3):389–397. [https://doi.](https://doi.org/10.1016/j.ygcen.2011.11.008) [org/10.1016/j.ygcen.2011.11.008](https://doi.org/10.1016/j.ygcen.2011.11.008)
- [Tsubota T, Takahashi Y, Kanagawa H.](#page-4-24) 1987. Changes in serum progesterone levels and growth of fetuses in Hokkaido brown bears. Bears: Their Biology and Management 7:355–358. [https://doi.](https://doi.org/10.2307/3872643) [org/10.2307/3872643](https://doi.org/10.2307/3872643)
- [Tyndale-Biscoe CH, Hearn JP, Renfree MB.](#page-4-25) 1974. Review control of reproduction in macropodid marsupials. Journal of Endocrinology 63(3):589–614. <https://doi.org/10.1677/joe.0.0630589>
- [Tyndale-Biscoe H, Renfree M.](#page-4-26) 1987. Ovarian function and control. In: Tyndale-Biscoe H, Renfree M, editors. Reproductive physiology of marsupials. Cambridge: Cambridge University Press; p. 203–257.
- [Upham NS, Esselstyn JA, Jetz W.](#page-1-26) 2019. Inferring the mammal tree: species-level sets of phylogenies for questions in ecology, evolution, and conservation. PLoS Biology 17(12):e3000494. [https://doi.](https://doi.org/10.1371/journal.pbio.3000494) [org/10.1371/journal.pbio.3000494](https://doi.org/10.1371/journal.pbio.3000494)
- [Urison NT, Buffenstein RB.](#page-0-28) 1995. Metabolic and body temperature changes during pregnancy and lactation in the naked mole rat (*Heterocephalus glaber*). Physiological Zoology 68(3):402–420. <https://doi.org/10.1086/physzool.68.3.30163776>
- [Van Raaij J, Schonk CM, Vermaat-Miedema SH, Peek M, Hautvast J.](#page-0-29) 1989. Body fat mass and basal metabolic rate in Dutch women before, during, and after pregnancy: a reappraisal of energy cost of pregnancy. The American Journal of Clinical Nutrition 49(5):765– 772. [https://doi.org/10.1093/ajcn/49.5.765](https://doi.org//10.1093/ajcn/49.5.765)
- [Verstegen-Onclin K, Verstegen J.](#page-0-30) 2008. Endocrinology of pregnancy in the dog: a review. Theriogenology 70(3):291–299. [https://doi.](https://doi.org/10.1016/j.theriogenology.2008.04.038) [org/10.1016/j.theriogenology.2008.04.038](https://doi.org/10.1016/j.theriogenology.2008.04.038)
- [Weitlauf HM, Greenwald GS.](#page-0-31) 1967. Maintenance of pregnancy in the hamster by a single corpus luteum from day 12. Journal of Reproduction and Fertility 14(3):489–491. [https://doi.org/10.1530/](https://doi.org/10.1530/jrf.0.0140489) [jrf.0.0140489](https://doi.org/10.1530/jrf.0.0140489)
- [Wiltbank MC.](#page-0-32) 1994. Cell types and hormonal mechanisms associated with mid-cycle corpus luteum function. Journal of Animal Science 72(7):1873–1883.<https://doi.org/10.2527/1994.7271873x>
- [Wu S-P, Li R, DeMayo FJ.](#page-0-33) 2018. Progesterone receptor regulation of uterine adaptation for pregnancy. Trends in Endocrinology and Metabolism 29(7):481–491. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.tem.2018.04.001) [tem.2018.04.001](https://doi.org/10.1016/j.tem.2018.04.001)
- [Wynne-Edwards K, Huck U, Lisk R.](#page-1-27) 1987. Infuence of pre-and postcopulatory pair contact on pregnancy success in Djungarian hamsters, *Phodopus campbelli*. Reproduction 80(1):241–249. [https://doi.](https://doi.org//10.1530/jrf.0.0800241) [org/10.1530/jrf.0.0800241](https://doi.org//10.1530/jrf.0.0800241)